

## Original Article

# Comparison of expression of CD1a and CD68 markers in skin leishmaniasis samples with positive and negative Leishman body

Fataneh Farokhpour<sup>1</sup>, Parvin Rajabi<sup>1</sup>, Bahare Abtahi Naeini<sup>2</sup>, Azar Naimi<sup>1</sup>

<sup>1</sup>Department of Pathology, School of Medicine, Isfahan University of Medical Sciences, Isfahan, Iran; <sup>2</sup>Department of Dermatology, School of Medicine, Isfahan University of Medical Sciences, Isfahan, Iran

Received November 4, 2020; Accepted July 7, 2021; Epub August 15, 2021; Published August 30, 2021

**Abstract:** Background: Leishmaniasis is one of the most important infectious illnesses around the world. Given the high commonness of this disease, specifically its skin type in Iran and due to the role of the Leishman bodies in diagnosis, the aim of present study was evaluating the expression of two CD1a and CD68 markers in cutaneous leishmaniasis lesions with and without Leishman bodies. Methods: In this case-control study, 70 skin samples of patients with cutaneous leishmaniasis (35 patients with Leishman body as case group and 35 patients without Leishman body as control group) were investigated during 2018-2019. The expression of CD1a and CD68 markers and immunohistochemistry staining (IHC) were investigated in this study. Results: The expression of CD1a in the group with Leishman body was significantly higher than group without Leishman body ( $P=0.01$ ), but there was no significant difference between groups as expression of CD68 ( $P=0.40$ ). The frequency of hyperkeratosis, parakeratosis, exocytosis, acanthosis, spongiosis, hydropic degeneration of basal cell layer, lichenoid reaction, pseudoepitheliomatous hyperplasia, ulcer, thinning of the epidermis, mononuclear cells, and extension of inflammation into lower dermis in the group with Leishman body was higher than group without Leishman body ( $P<0.05$ ). Conclusion: The expression of CD1a and other morphological findings help to diagnose the difference between leishmaniasis with and without Leishman body.

**Keywords:** CD1a, CD68, leishmaniasis, immunohistochemistry staining, morphological

## Introduction

Leishmaniasis is one of the six most remarkable infectious diseases around the world. Almost 350 million people are exposed to leishmaniasis and every 2 million new cases are being added to it continuously [1-3]. Clinical types observed in Iran include dry-type cutaneous leishmaniasis [4], wet type cutaneous leishmaniasis [5], Lymphadenitis [6] and visceral leishmaniasis [7]. In histopathology of cutaneous leishmaniasis, which is the most prevalent form of skin involvement, the dense and diffuse infiltration of histiocytic with lymphocytes and low plasma cells can be seen. Eosinophils and neutrophils are very low. The cytoplasm of the histiocytic is full of Lyman bodies and when these bodies are abundant, they can also be seen out of the cells [8-10]. In

several studies, importance of the immunohistochemistry of inflammatory cells in the cutaneous lesions of different species of leishmaniasis were investigated for diagnosis of cutaneous leishmaniasis [11, 12]. Based on these studies, it has been found that cellular immunity has an important role in the pathogenesis of cutaneous leishmaniasis. T-lymphocytes are the major factor in lymphocytic infiltration and the control of parasite replication in the cutaneous leishmaniasis [13-16].

It has been observed that the molecules of CD1, group one, including human CD1a, CD1b and CD1c, lipid-mycolipids and glycolipids to specialized T-cells. This can assuredly help antimicrobial resistance with the production of interferon-gamma (IFN- $\gamma$ ) [17-19] and the antimicrobial protein of granulation [20, 21].

Therefore, CD1 group I protein probably participates in the protected immune response against complex intracellular pathogens such as Leishman species known as glycolipid antigens. On the other hand, the expression of CD1a, CD1b, CD1c, and CD1d has been seen more in CD68 + cells and these cells can present external antigens to T cells, while the normal arterial samples do not express CD1 molecules [19, 22]. In fact, leishmaniasis parasites interact with the types of host T cells and infect them; CD68 + macrophages and dendritic cell 1a (DC1a) are the most important cells that adjust the outcome of infection. After the first absorption of amastigotes by macrophages into phagosome, subsequent fusion will happen with the lysosome, and parasites must stay alive in this environment. This is probably one of the most controversial environments for many diseases, and the leishmaniasis is one the protozoa that can survive and reproduce under difficult circumstances. Knowing how these organisms can survive and manipulate host cells for the purpose of replication and transfer is quite critical to the design of new drugs or treatment strategies against the disease [23, 24]. In this regard, Taheri and colleagues in their study (2017) showed that chronic cutaneous leishmaniasis had a significant relationship with macrophage CD68 + and dendritic CD1a + cells and macrophage + CD68 and CD1A + increases the duration of chronic leishmaniasis [25]. Correlation of CD1a and CD68 in patients with leishmaniasis was evaluated in the previous studies and indicated expression of these markers could be increase in the chronic leishmaniasis [25-28]. However, in the present study, the difference between leishmaniasis with and without Leishman body was compared for first one in the Iran. Therefore, due to the high prevalence of this disease, the role of the Leishman bodies in diagnosis, considering the assumption that the samples of skin leishmaniasis in the case with or without the Leishman body are similar in immunohistochemistry staining (IHC) staining and have no difference with each other, so it can be utilized instead of polymerase chain reaction (PCR), and in addition to the lack of study in this field, this study examined expression of two CD1a and CD68 markers in cutaneous leishmaniasis lesions with and without Leishman bodies.

### Materials and methods

In case-control group, 70 skin samples of patients with cutaneous leishmaniasis were entered in study between 2018-2019. The current study was approved in ethical committee of Isfahan University of Medical Sciences (IR.MUI.MED.REC.1398.176) and all patients signed the informed consent to for participation in the study.

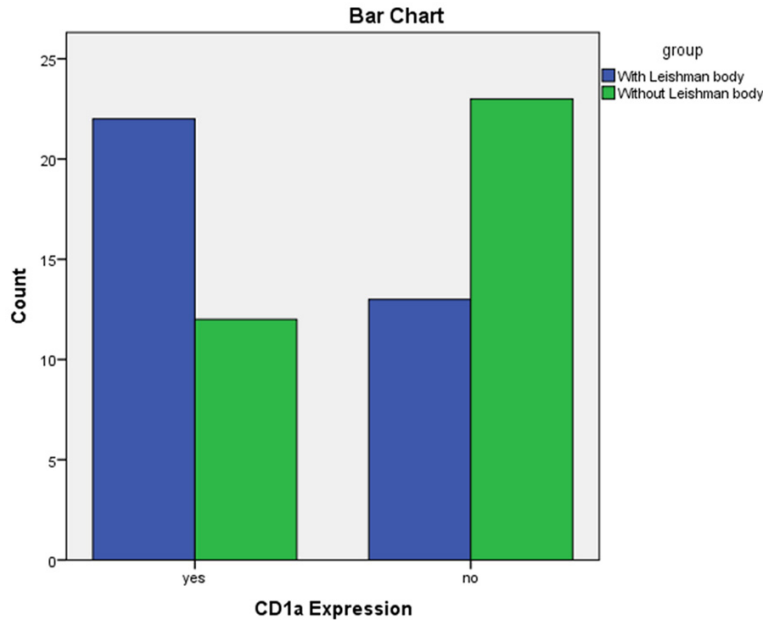
Among 70 patients, including 35 cases with Leishman body (seen in their pathology culture in paraffin blocks) were considered case group and 35 cases without Leishman body (not seen in their pathology culture but PCR was confirmed leishmaniasis) were considered control group. Therefore, the present study was conducted on two groups of patients with cutaneous leishmaniasis with and without Leishman body in their skin samples pathology. All patients were selected in Isfahan, Iran, and demographic information of patients including results, age, sex, location, and duration of were collected.

The treatment of the patients with lesions were based on age, duration of disease, stage of disease, and place of lesion, that dermatologist recommended different treatment including [29] cryotherapy, infiltration of sodium stibogluconate at 0.3-0.8 mL, local heat therapy at 40-42°C, and topical paromomycin.

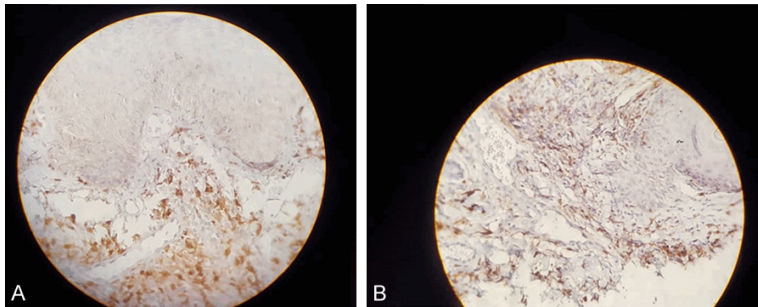
### *Histopathology changes*

The paraffin block cuts of the biopsy, which were fixed with formalin 10% (neutral buffered formalin), were stained with Hematoxylin and eosin and investigated by light microscopy. In addition to topographic descriptions, epidermis and dermis changes were recorded. Epidermis changes may include hyperkeratosis, Parakeratosis, exocytosis, acanthosis, spongiosis, and hydropic degeneration of basal cell layer, lichenoid reaction, pseudoepitheliomatous hyperplasia, psoriasiform epidermal hyperplasia, Ulcer, and thinning of the epidermis. Dermal changes involved severity and extension of inflammation (upper, middle and lower of derma) as well as the type and amount of inflammatory cells including lymphocyte, plasma cell, histiocyte, neutrophils, and also presence or absence of granuloma, necrosis, fibrosis and abscess.

# Skin leishmaniasis



**Figure 1.** CD1a expression based on groups as with or without Leishman body (Independent T test).



**Figure 2.** Microscopic findings of CD1a and CD68 in immune cells, A: CD1a in dendritic cells (x40), B: CD68 in macrophage (x40).

## Immunohistochemistry changes

IHC, histologic 3-micron slices were put on the special Silanization and paraffin trouble was performed with xylene, ethanol and water. Antigen Retrieval was performed with (pH=9) Tris-EDTA buffer and microwave heat during 10 minutes. The activity of peroxidase with 0.5% methanol solution and  $H_2O_2$  block were performed and incubated with key special antibodies in certain concentration of wet environment and room temperature during 30 minutes.

The characteristics of used monoclonal antibodies were CD1a (DAKO M3571), dilution factor {df}: 1/50 and CD68 (DAKO M0814, df: 1/100). The EnVision was performed to solu-

tion that was contains secondary antibody bonded to biotin and streptavidin bonded to peroxidase. DAB chromogene was used to determine the anti-antibody bands and the fields were stained with Hematoxylin (**Figure 2**).

All in all, the collected data was analyzed using Statistical Package for Social Sciences (SPSS) software version 24 (IBM, USA), and data were shown based on mean and standard deviation or frequency and percentage. Also Fisher exact test, chi-square, the independent T-Test and Pearson correlation coefficient were used to compare variables between two groups. The significance level was considered less than 0.05.

## Results

### Demographical

Patients were categorized into two groups as leishmaniasis with (24 male and 11 female) and without (27 male and 8 female) Leishman body; there was no significant differences between groups in terms of

age, gender, region of lesions, and duration of disease ( $P>0.05$ ).

### Expression of markers

The expression of CD1a in the group with Leishman body was significantly higher than group without Leishman body ( $P=0.01$ ), but there was no significant difference between groups in terms of the expression of CD68 ( $P=0.40$ ) (**Figure 1**; **Table 1**).

### Morphological characteristics

According to morphological characteristics, the frequency of hyperkeratosis, parakeratosis, exocytosis, acanthosis, spongiosis, hydropic

## Skin leishmaniasis

**Table 1.** Variables of study between two groups

Variables		With Leishman body	Without Leishman body	P-value
Age (year) f (mean ± SD)		29.48±10.51	32.74±10.58	0.20
Gender (m/f)		24/11	27/8	0.29
Region	Lower limb	20 (57.1%)	24 (68.6%)	0.40
	Upper limb	12 (34.3%)	7 (20%)	
	Others	3 (8.6%)	4 (11.4%)	
Duration of disease [33] (mean ± SD)		15.51±4.13	15.20±3.04	0.71
Expression of CD1a		22 (62.9%)	12 (34.3%)	0.01
Expression of CD68		19 (54.3%)	17 (48.6%)	0.40

**Table 2.** Morphological characteristics of lesions in the groups with and without Leishman body

Morphological characteristics		With Leishman body	Without Leishman body	P-value*	
Hyperkeratosis		33 (94.3%)	15 (42.9%)	<0.001	
Parakeratosis		26 (74.3%)	6 (17.1%)	<0.001	
Exocytosis		24 (68.6%)	11 (31.4%)	0.002	
Acanthosis		22 (62.9 %)	10 (28.6%)	0.004	
Spongiosis		26 (74.3%)	8 (22.9%)	<0.001	
hydropic degeneration of basal cell layer		34 (97.1%)	13 (37.1%)	<0.001	
lichenoid reaction		30 (85.7%)	19 (54.3%)	0.004	
pseudoepitheliomatous hyperplasia		28 (80%)	8 (22.9%)	<0.001	
Psoriasiform epidermal hyperplasia		18 (51.4%)	13 (37.1%)	0.16	
Ulcer		35 (100%)	3 (8.6%)	<0.001	
Thinning of the epidermis		35 (100%)	13 (37.1%)	<0.001	
Epithelial cell	PMN	<10	11 (31.4%)	0.13	
		Absence	24 (68.6%)		
	Eosinophils	<10	6 (17.1%)	0.23	
		Absence	29 (82.9%)		
	Mononuclear	>10	28 (80%)	1 (2.9%)	<0.001
		<10	7 (20%)	20 (57.1%)	
Absence		0	14 (40%)		
The extent of the inflammation	Upper of dermis	1 (2.9%)	18 (51.4%)	<0.001	
	Middle of dermis	12 (34.3%)	16 (45.7%)		
	Lower of dermis	22 (62.9%)	1 (2.9%)		
Granuloma		2 (5.7%)	0	0.24	
Necrosis		4 (11.4%)	0	0.05	
Fibrosis		4 (11.4%)	0	0.05	

PMN: Polymorphonuclear neutrophil, \*Chi Square.

degeneration of basal cell layer, lichenoid reaction, pseudoepitheliomatous hyperplasia, ulcer, thinning of the epidermis, mononuclear cells, and extension of inflammation into a lower dermis in the group with Leishman body was higher than group without Leishman body ( $P<0.05$ ); but there was no significant difference between groups in terms of psoriasiform epidermal hyperplasia, PMNs and eosinophils cells, granuloma, necrosis and fibrosis of lesions ( $P>0.05$ ) (**Table 2**).

### Discussion

According to the results of this study, the expression of CD1a was significantly higher in patients with Leishman body, but there was no difference between patients with and without Leishman bodies in CD68 expression. Therefore, CD1a marker as a marker in determining the Leishman body can be effective; on the other hand, patients who had a Leishman body and patients without it were different in mor-

## Skin leishmaniasis

phological characteristics. In terms of the frequency of hyperkeratosis, parakeratosis, exocytosis, acanthosis, spongiosis, hydropic degeneration of basal cell layer, lichenoid reaction, pseudoepitheliomatous hyperplasia, and ulcer, thinning of the epidermis, mononuclear cells, and extension of inflammation in patients with Leishman body was significantly more compared with patients without Leishman body.

In this regard, Taheri and colleagues (2017) showed that chronic cutaneous leishmaniasis had a significant relationship with macrophage CD68 + and dendritic CD1a + cells and macrophage + CD68 and CD1A + increases the duration of chronic leishmaniasis [25].

Karram et al. in 2012, showed how the intra-epidermal amastigotes express CD1a. They examined the removal of epidermal leishmaniasis in 60 of 212 biopsy samples from cutaneous leishmaniasis. In all cases, the positive CD1a in the intra-epidermal amastigotes was determined. In this study, authors indicated that low level CD1a was associated high risk of leishmaniasis. Karram and his colleagues provided two plausible explanations for this abnormal safety phenotype of amastigotes [1]: the amastigotes had gained CD1a after leaving the dendritic cells that expressed this marker with the exocytosis process; Or [2]: the leishmaniasis level has shown a mutual reaction with CD1a antibodies (which is less than the authors' opinion [26]. Jabbour et al. (2015) also examined 11 skin biopsy samples from 33 patients with *L. tropica* or major *L.* and added new data about the process of acquiring CD1a by leishmaniasis. The cultured amastigotes were CD1a. Therefore, the amastigotes will gain CD1a during the host infection [27]. In the present study, there was a significant relationship between expression of CD1a and leishmaniasis, which was higher in patients with Leishman body.

Based on Karram et al. and the report of Jabbour et al., the authors studied the items created by *Tropica L.* and Major *L.* None of these species is found in our area, whereas the cases of leishmaniasis are created exclusively by the *infantum L.* However, there are some major differences between the 3-species genome, including the genome size, guanine and cytosine content, and the number of genes.

A recent study by Dias-Polak et al. in 2017 showed that properties of skin lesions in leishmaniasis could be varied. They also reported that the presence of Leishman body or absence of it might influence these varieties [28]. Furthermore, in another study by Meymandi and others in 2009, it was shown that the presence of Leishman body could make differences in histological and immunological details related to the disease including CD1a [12]. These results are in line with the findings of our study. Herein we showed that expression of CD1a is significantly different among skin samples with or without Leishman body. Our results highlighted higher expression of CD1a in patients with Leishman body in their skin samples.

Another study was carried out by Tabrizchi et al. and they focused on histopathological properties of skin lesions in leishmaniasis and assessed these features by the means of IHC and reported that CD1a has higher expression rate in patients with Leishman body. They also showed no significant differences between samples concerning on other markers including CD68 [30]. The results of Tabrizchi study was in line with our results.

Moreover, histological and morphological evaluation of leishmaniasis lesions was performed by Asgari and others in 2007 in Iran. They noticed increased frequency of characteristics including exocytosis, acanthosis, spongiosis, hydropic degeneration of basal cell layer and lichenoid reaction in patients with Leishman body in their skin lesions [31]. These results are also in line with the findings of Oryan and colleagues in 2008 [32] and also in line with our findings.

As it was mentioned before, we depicted that histological characteristics such as hyperkeratosis, parakeratosis, exocytosis, acanthosis, spongiosis, hydropic degeneration of basal cell layer, lichenoid reaction, pseudoepitheliomatous hyperplasia, ulcer, thinning of the epidermis, mononuclear cells, and extension of inflammation were more common in patients who had Leishman body in their skin samples. Actually, these results could be helpful in diagnosis and treatments of patients with leishmaniasis. Utilization of immunological markers and their associations with pathogenesis of the

disease could be helpful in further novel treatments.

## Disclosure of conflict of interest

None.

**Address correspondence to:** Fataneh Farokhpour, Department of Pathology, School of Medicine, Isfahan University of Medical Sciences, Isfahan 8174673461, Iran. Tel: +98 913 726 6066; E-mail: fatanehfarkh@gmail.com

## References

- [1] Desjeux P. Leishmaniasis: current situation and new perspectives. *Comp Immunol Microbiol Infect Dis* 2004; 27: 305-318.
- [2] Noazin S, Khamesipour A, Moulton LH, Tanner M, Nasser K, Modabber F, Sharifi I, Khalil EA, Bernal ID, Antunes CM and Smith PG. Efficacy of killed whole-parasite vaccines in the prevention of leishmaniasis-a meta-analysis. *Vaccine* 2009; 27: 4747-4753.
- [3] Haddad S, Ghadimi K, Abrishamkar R and Asl NSM. Comparing laparoscopy and laparotomy procedures in the radical hysterectomy surgery for endometrial cancer: a basic review. *Am J Transl Res* 2021; 13: 2456-2461.
- [4] Momeni A and Aminjavaheri M. Clinical picture of cutaneous leishmaniasis in Isfahan, Iran. *Int J Dermatol* 1994; 33: 260-265.
- [5] Yaghoobi-Ershadi M and Javadian E. Zoonotic cutaneous leishmaniasis to the north of Isfahan. Human infection in 1991. *Bull Soc Pathol Exot* 1995; 88: 42-45.
- [6] Azadeh B. "Localized" leishmania lymphadenitis: a light and electron microscopic study. *Am J Trop Med Hyg* 1985; 34: 447-455.
- [7] Shamsi S, Eslammanesh T, Dabiri S, SHasi Meimandi M and Naji M. The histopathological changes and immunohistochemical findings of acute, chronic nonlupoid and chronic lupoid types of cutaneous leishmaniasis. *Journal of Kerman University of Medical Sciences* 2010; 17: 281-296.
- [8] Choi CM and Lerner EA. Leishmaniasis as an emerging infection. *J Invest Dermatol Symp Proc* 2001; 6: 175-82.
- [9] Lehoczky L, Southworth AB, Martinez GZ, Belfort MA, Shamshirsaz AA, Shamshirsaz A, Sanz Cortes M, Nassr AA, Donepudi R, Whitehead WE, Johnson R, Meshinchi N and Espinoza J. Magnesium sulfate titration reduces maternal complications following fetoscopic closure of spina bifida. *Prenat Diagn* 2021; [Epub ahead of print].
- [10] Espinoza J, Shamshirsaz AA, Sanz Cortes M, Pammi M, Nassr AA, Donepudi R, Whitehead WE, Castillo J, Johnson R, Meshinchi N, Sun R, Krispin E, Corroenne R, Lee TC, Keswani SG, King A and Belfort MA. Two-port, exteriorized uterus, fetoscopic meningocele closure has fewer adverse neonatal outcomes than open hysterotomy closure. *Am J Obstet Gynecol* 2021; [Epub ahead of print].
- [11] Morgado F, Schubach A, Rosalino C, Quintella L, Santos G, Salgueiro M and Conceição-Silva F. Is the in situ inflammatory reaction an important tool to understand the cellular immune response in American tegumentary leishmaniasis? *Br J Dermatol* 2008; 158: 50-8.
- [12] Meymandi S, Dabiri S, Shamsi-Meymandi M, Nikpour H and Kharazmi A. Immunophenotypic pattern and cytokine profiles of dry type cutaneous leishmaniasis. *Arch Iran Med* 2009; 12: 371-6.
- [13] Gimblet C, Loesche MA, Carvalho L, Carvalho EM, Grice EA, Artis D and Scott P. IL-22 protects against tissue damage during cutaneous leishmaniasis. *PLoS One* 2015; 10: e0134698.
- [14] Bangert C, Brunner PM and Stingl G. Immune functions of the skin. *Clin Dermatol* 2011; 29: 360-376.
- [15] Zadeh AR, Askari M, Azadani NN, Ataei A, Ghadimi K, Tavosi N and Falahatian M. Mechanism and adverse effects of multiple sclerosis drugs: a review article. Part 1. *Int J Physiol Pathophysiol Pharmacol* 2019; 11: 95-104.
- [16] Zadeh AR, Ghadimi K, Ataei A, Askari M, Sheikhhinia N, Tavosi N and Falahatian M. Mechanism and adverse effects of multiple sclerosis drugs: a review article. Part 2. *Int J Physiol Pathophysiol Pharmacol* 2019; 11: 105-114.
- [17] Sieling PA, Chatterjee D, Porcelli SA, Prigozy TI, Mazzaccaro RJ, Soriano T, Bloom BR, Brenner MB, Kronenberg M, Brennan PJ, et al. CD1-restricted T cell recognition of microbial lipoglycan antigens. *Science* 1995; 269: 227-230.
- [18] Etemadifar M, Ghadimi M, Ghadimi K and Alshehbosoul F. The serum amyloid  $\beta$  level in multiple sclerosis: a case-control study. *Neurol Sci* 2017; 3: 214-221.
- [19] Ashtari F, Madanian R, Shaygannejad V, Zarkesh SH and Ghadimi K. Serum levels of IL-6 and IL-17 in multiple sclerosis, neuromyelitis optica patients and healthy subjects. *Int J Physiol Pathophysiol Pharmacol* 2019; 11: 267.
- [20] Stenger S, Hanson DA, Teitelbaum R, Dewan P, Niazi KR, Froelich CJ, Ganz T, Thoma-Uszynski S, Melián A, Bogdan C, Porcelli SA, Bloom BR, Krensky AM and Modlin RL. An antimicrobial activity of cytolytic T cells mediated by granulysin. *Science* 1998; 282: 121-125.
- [21] Farrokhi M, Dabirzadeh M, Dastravan N, Etemadifar M, Ghadimi K, Saadatpour Z and

## Skin leishmaniasis

- Rezaei A. Mannose-binding lectin mediated complement pathway in autoimmune neurological disorders. *Int J Physiol Pathophysiol Pharmacol* 2016; 15: 251-256.
- [22] Behar S and Porcelli SA. CD1-restricted T cells in host defense to infectious diseases. In: editors. *T cell activation by CD1 and lipid antigens*. Springer; 2007. pp. 215-250.
- [23] Xin L, Li K and Soong L. Down-regulation of dendritic cell signaling pathways by *Leishmania amazonensis* amastigotes. *Mol Immunol* 2008; 45: 3371-3382.
- [24] Isnard A, Shio MT and Olivier M. Impact of *Leishmania* metalloprotease GP63 on macrophage signaling. *Front Cell Infect Microbiol* 2012; 2: 72.
- [25] Taheri E, Dabiri S, Meymandi MS and Saedi E. Possible interrelationship of inflammatory cells in dry type cutaneous leishmaniasis. *Iran J Pathol* 2017; 12: 119.
- [26] Fernandez-Flores A and Rodriguez-Peralto JL. Morphological and immunohistochemical clues for the diagnosis of cutaneous leishmaniasis and the interpretation of CD1a status. *J Am Acad Dermatol* 2016; 74: 536-543.
- [27] Jabbour M, Issa G, Charafeddine K, Simaan Y, Karam M, Khalifeh H, Habib R and Khalifeh I. The immune microenvironment in cutaneous leishmaniasis. *J Eur Acad Dermatol Venereol* 2015; 29: 1170-1179.
- [28] Dias-Polak D, Geffen Y, Ben-Izhak O and Bergman R. The role of histopathology and immunohistochemistry in the diagnosis of cutaneous leishmaniasis without “discernible” leishman-donovan bodies. *Am J Dermatopathol* 2017; 39: 890-895.
- [29] At:<http://www.cdc.gov/nceh/lead/publications/leadandpregnancy2010.pdf>. Retrieved March 7, 2012. CfDCaPGftiamoleipalwAGCA.
- [30] Tabrizchi H, Dabiri S, Soutodehnejad A, Azadeh B and Malcolm MMH. Localized leishmania lymphadenitis: a morphologic and immunohistochemical study. *Iran J Pathol* 2007; 2: 41-44.
- [31] Asgari Q, Motazedian MH, Mehrabani D, Oryan A, Hatam GR, Owji SM and Paykari H. Zoonotic cutaneous leishmaniasis in Shiraz, Southern Iran: a molecular, isoenzyme and morphologic approach. *J Res Med Sci* 2007; 12: 7-15.
- [32] Oryan A, Mehrabani D, Owji S, Motazedian M, Hatam G and Asgari Q. Morphologic changes due to cutaneous leishmaniasis in BALB/c mice experimentally infected with *leishmania major*. *J Appl Behav Anal* 2008; 34: 87-92.
- [33] Battleday RM and Brem AK. Modafinil for cognitive neuroenhancement in healthy non-sleep-deprived subjects: a systematic review. *Eur Neuropsychopharmacol* 2015; 25: 1865-1881.